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Supplemental Information

Individual Identifiability Predicts

Population Identifiability

in Forensic Microsatellite Markers

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Figure S1. Properties of DAPC inferences for *K*=4 **clusters.** (A-D) DAPC inferences. Each sampled individual is represented by a vertical line. Colors represent clusters, and the length of the line segment displayed in a color is proportional to the estimated membership for the associated cluster. (A) CODIS loci. $B_{DAPC}=0.868$, $S_{DAPC}=0.684$, $T_{DAPC}=0.838$. (B) 13-locus null dataset. $B_{DAPC}=0.548$, $S_{DAPC}=0.479$, $T_{DAPC}=0.374$. (C) 13 random non-CODIS tetranucleotide markers. $B_{DAPC}=0.867$, $S_{DAPC}=0.677$, $T_{DAPC}=0.841$. (D) 779 non-CODIS loci. $B_{DAPC}=1.000$, $S_{DAPC}=1.000$. For (B) and (C), the solution shown has the median *S* among 1000 runs. (E-G) Distributions of three indices describing DAPC inferences. The value for the CODIS loci (dashed lines) appears together with the distributions for 1000 solutions using random sets of 13 non-CODIS loci (histogram). (E) Clusteredness B_{DAPC} . (F) Similarity-to-full-data S_{DAPC} . (G) Sorting accuracy T_{DAPC} . The percentile for the CODIS loci is 54.1 for B_{DAPC} , 60.7 for S_{DAPC} , and 64.4 for T_{DAPC} . The figure design follows Figure 2.

Table S1. *B* and *S* percentiles of the median STRUCTURE replicate using the CODIS loci in relation to the distribution from runs with 1000 random sets of 13 non-CODIS loci. The *K*=4 median-*S* replicate is plotted in Figure 2A.

K	Clusteredness B	Similarity S
2	36.8	48.8
3	46.6	12.1
4	53.4	51.8
5	49.5	45.7
6	38.3	42.4

Table S2. Mean confusion matrices sorting 978 individuals into K=4 **clusters using the CODIS loci.** For the top (STRUCTURE-based) matrix, the value of *T* is 74%; the median *T* is 75% across 1000 random 13-locus sets and 27% across 1000 null datasets. For the bottom (DAPC-based) matrix, the value of T_{DAPC} is 84%; the median T_{DAPC} is 82% across 1000 random 13-locus sets and 36% across 1000 null datasets. The matrices summarize STRUCTURE and DAPC inferences of the form plotted in Figures 2A-2D and S1A-S1D.

	ST	RUCTURE-ba	sed confusio	n matrix		
	Cluster 1	Cluster 2	Cluster 3	Cluster 4	Total	Sorting
	(orange)	(blue)	(pink)	(violet)		accuracy (%)
Africa	85.4	2.7	3.1	2.8	94	90.9
Western Eurasia	101.5	275.3	78.7	76.6	532	51.7
East Asia/Pacific	29.9	16.8	172.7	49.6	269	64.2
America	1.6	1.6	5.9	73.8	83	88.9
		DAPC-based	l confusion r	natrix		
	Cluster 1	Cluster 2	Cluster 3	Cluster 4	Total	Sorting
	(orange)	(blue)	(pink)	(violet)		accuracy (%)
Africa	72	20	2	0	94	76.6
Western Eurasia	5	490	35	2	532	92.1
East Asia/Pacific	1	54	211	3	269	78.4
America	0	7	3	73	83	88.0

Table S3. $\overline{F_{ST}}$ -adjusted correlations between measures of individual identifiability and population identifiability for 1000 random sets of 13 non-CODIS loci. For convenience, the correlation between \overline{H} and \overline{M} is copied from the top (STRUCTURE) to the bottom (DAPC) part of the table. The partial correlations adjust the correlations that appear in Figure 3 and Table S4 for $\overline{F_{ST}}$.

	Dive	rsity measures	STRUC of ances	TURE-base stry inform	ed measures nation at <i>K</i> =4
	\overline{H}	\overline{M}	В	S	Т
\overline{H}		-0.97	0.51	0.36	0.21
\overline{M}			-0.55	-0.39	-0.23
В				0.75	0.58
S					0.89
	Dive	rsity measures	DAPC-based measures of		
			ancest	ry informa	tion at <i>K</i> =4
	\overline{H}	\overline{M}	B _{DAPC}	S_{DAPC}	$T_{\rm DAPC}$
\overline{H}		-0.97	0.44	0.47	0.39
\overline{M}			-0.45	-0.48	-0.41
$B_{\rm DAPC}$				0.91	0.74
$S_{\rm DAPC}$					0.88

Table S4. Correlations between measures of individual identifiability and DAPC-based measures of population identifiability for 1000 random sets of 13 non-CODIS loci. The table reports analogous correlations to those that appear in Figure 3, and for convenience, it copies from that figure the correlations between \overline{H} , \overline{M} , and $\overline{F_{ST}}$.

		Diversity measures	Variance partition	DAP ancest	C-based meas ry information	ures of n at <i>K</i> =4
	\overline{H}	\overline{M}	$\overline{F_{ST}}$	B _{DAPC}	$S_{\rm DAPC}$	$T_{\rm DAPC}$
\overline{H}		-0.97**	-0.14*	0.28**	0.29**	0.23**
\overline{M}			0.11*	-0.31**	-0.31**	-0.26**
$\overline{F_{ST}}$				0.56**	0.60**	0.58**
B _{DAPC}					0.94**	0.83**
S _{DAPC}						0.92**
$\frac{S_{\text{DAPC}}}{*p < 0.05}$.						0.72

***p*<0.001.

Table S5. Correlations between STRUCTURE-based and DAPC-based measures of population identifiability for 1000 random sets of 13 non-CODIS loci. The table provides correlations of the values that appear in the histograms in Figures 2E-2G and S1E-S1G.

	STRUCTURE-based measures		DAPC-based measures			
	of ancestry information at K=4			of ancestry information at K=4		
	В	S	Т	B_{DAPC}	S_{DAPC}	T_{DAPC}
В		0.87	0.71	0.70	0.73	0.71
S			0.91	0.67	0.69	0.67
Т				0.51	0.54	0.57
B_{DAPC}					0.94	0.83
SDAPC						0.92

Supplemental Experimental Procedures

Data

Samples were drawn from the Human Genome Diversity Panel (HGDP-CEPH) H1048 subset [S1]. In classifying individuals as in past studies [S2-S6], the sample set included 94 people from Sub-Saharan Africa, 155 from Europe, 206 from Central and South Asia, 171 from the Middle East, 234 from East Asia, 35 from Oceania, and 83 from the Americas.

We combined 783 previously studied microsatellites [S4] with the 13 autosomal CODIS loci; CODIS genotypes were produced by Bode Technology Group (Lorton, VA) using the Promega PowerPlex 16 HS System. Four loci appeared in both marker sets; we discarded them from the data of [S4]. Locus-wise missing data ranged from 0% to 12.2% (mean 3.6% across the 792 loci). CODIS loci had no missing data.

Note that the non-CODIS loci are comparable to the CODIS loci in being highly polymorphic, widely spaced, and generally non-genic [S7]. Even the more closely spaced among them are generally pairwise-independent, producing negligible linkage disequilibrium in unstructured populations [S8].

Heterozygosity H

We computed locus-specific heterozygosity using the standard unbiased estimator [S9]

$$H = \frac{2N(1 - \sum_{j=1}^{J} p_j^2)}{2N - 1},\tag{1}$$

where *N* is the diploid sample size, *J* is the number of alleles at the locus, and p_j is the frequency of allele *j*. Allele frequencies were estimated excluding individuals with missing data. For locus sets, we evaluated \overline{H} , the mean *H* across loci.

Match probability M

We calculated the locus-specific diploid random match probability as [S10,S11]

$$M = 2\left(\sum_{j=1}^{J} p_j^2\right)^2 - \sum_{j=1}^{J} p_j^4.$$
 (2)

For sets of loci, we computed \overline{M} , the geometric mean of M. The product of M values across loci directly measures a random match probability for a panel; the geometric rather than arithmetic mean reflects the contribution of a typical locus.

Fst

We computed F_{ST} as the sum of among-region and among-population-within-region variance components in a threelevel hierarchical partition. This partition, performed with the estimators of [S9] at each locus, considered individuals as distributed across the seven geographic regions. $\overline{F_{ST}}$ values for locus sets were obtained by averaging estimates across loci.

STRUCTURE

We conducted unsupervised model-based clustering using STRUCTURE 2.3.4 [S12,S13], employing an admixture model with correlated allele frequencies in 20,000 steps, 10,000 of which were allocated to burn-in. We set parameters α and λ to 1.

To produce "null" datasets with no ancestry information, we permuted alleles at each of the CODIS loci independently, not preserving co-occurrences of allele pairs within individuals or co-occurrences within individuals of genotypes across loci. This procedure amounts to applying Hardy-Weinberg and linkage equilibrium, holding allele frequencies constant.

K=4 was the largest *K* for which, in CODIS analyses, each cluster consistently had individuals for which membership in the cluster exceeded that in other clusters. *K*=4 was also inferred with the ΔK [S14] ($\Delta K(4)$ =3.1; next was $\Delta K(3)$ =1.6) and Pr(*K*=*k*|*X*) [S12] statistics (Pr(*K*=4|*X*)≈1). The latter measure was computed using the median Pr(*X*|*K*=*k*) from 1000 runs, employing CLUMPAK [S15]. STRUCTURE solutions were plotted using DISTRUCT [S16], considering the "median" run for the purpose of plotting as the 500th smallest value among 1000.

Clusteredness B

Given *I* individuals and *K* clusters, STRUCTURE estimates an $I \times K$ matrix of membership coefficients. The membership for individual *i* in cluster *k* is q_{ik} . *B* is calculated from a STRUCTURE solution without reference to other solutions [S4]:

$$B = \frac{1}{I} \sum_{i=1}^{I} \sqrt{\frac{K}{K-1} \sum_{k=1}^{K} (q_{ik} - 1/K)^2}.$$
(3)

Similarity S

Following [S17], the similarity S measures concordance of a target STRUCTURE solution with comparison solutions at the same K. Let $Q_1, Q_2, ..., Q_L$ be membership matrices from STRUCTURE runs on the full dataset. Using G' for the similarity measure in eq. 6 of [S17], the similarity to the comparison matrices of a target matrix R of the same dimensions is

$$S(R) = \frac{1}{L} \sum_{l=1}^{L} G'(Q_l^P, R).$$
(4)

The superscript *P* indicates that columns of Q_1 have been permuted in the way that maximizes similarity between Q_1 and *R* [S17].

Sorting accuracy T

The sorting accuracy T measures the extent to which STRUCTURE placed individuals in clusters together with other members of the same geographic region. For each of the 100 STRUCTURE replicates at K=4 with all 779 non-CODIS loci, the cluster with the largest membership coefficient was identified for each individual. Next, we determined which of the seven geographic regions "co-clustered," where two regions co-cluster if pluralities of individuals belonging separately to the two regions sort into the same cluster. The same co-clustering pattern occurred in all 100 replicates, producing four super-regions.

For each STRUCTURE solution estimated using the CODIS markers or 13 random loci, we constructed a confusion matrix W, where $W_{i,j}$ is the number of individuals from super-region *i* placed into cluster *j*. We associated each cluster bijectively with one of the four super-regions by permuting columns of W to maximize the number of correctly classified individuals, $\sum_{i=1}^{4} W_{i,i}$. Then

$$T = \frac{1}{4} \sum_{i=1}^{4} \frac{W_{i,i}}{\sum_{j=1}^{4} W_{i,j}}.$$
(5)

T is the mean frequency with which individuals in a super-region sort into the cluster associated with that super-region.

Correlations

We assessed significance of correlations involving H, M, and F_{ST} by permutation, separately permuting locus-wise values of H, M, and F_{ST} appearing in the 1000 sets of 13 random markers. After applying the permutation, we recomputed \overline{H} , \overline{M} , $\overline{F_{ST}}$, and the Pearson correlations for the 1000 sets. We examined permutation distributions with 10,000 permutations. In comparisons of two among \overline{H} , \overline{M} , and $\overline{F_{ST}}$, independently permuted values were used for both statistics. Because B, S, and T are not derived from locus-level information, for significance tests involving two among B, S, and T, we instead compared correlations to a $N(0, 0.06^2)$ distribution. This distribution was chosen conservatively by noting that permutation distributions of correlations with \overline{H} , \overline{M} , and $\overline{F_{ST}}$ were roughly normal with means near 0 and smaller standard deviations, 0.048 to 0.057.

To compute partial correlations, we regressed \overline{H} , \overline{M} , B, S, and T for each set of 13 markers on $\overline{F_{ST}}$. We then evaluated Pearson correlations of residuals from these five least-squares linear regressions.

Principal component analysis

We used principal component analysis (PCA) to produce population structure estimates in a parallel manner to that used in our STRUCTURE analysis. For PCA with multiallelic markers, we placed each distinct allelic type in its own column in the input data matrix [S18]. In this matrix, each individual was represented by a row, and the entry for a row and column was the number of copies of an allelic type present in the associated individual. Missing entries were imputed as the column mean of non-missing values [S19]. Prior to running PCA, we centered and standardized columns to have mean 0 and variance 1 [S20,S21].

To facilitate comparison with STRUCTURE, we post-processed PCA output using linear discriminant analysis (LDA), as applied for population genetics in the <u>discriminant analysis</u> of <u>principal components</u> approach (DAPC [S19]). This method converts each vector of principal components (PCs) representing an individual into a vector of membership probabilities in clusters estimated by LDA from the set of individual PC vectors. Prior to implementing LDA, we performed dimensionality reduction on PC vectors, considering only the PCs associated with the largest eigenvalues required for obtaining 90% of the variance in the initial data matrix [S22]. In parallel to our STRUCTURE analysis, we applied LDA to produce membership probabilities in *K*=4 clusters. We implemented our PCA and LDA analyses using the ADEGENET package in R [S23,S24].

We applied DAPC to the CODIS set, the non-CODIS set, 1000 null datasets, and the same 1000 random non-CODIS datasets used in the STRUCTURE analysis. Because the procedure is deterministic—unlike in the stochastic STRUCTURE analysis—we performed only one replicate DAPC with each marker set.

The matrix of membership probabilities estimated with DAPC, containing an estimated membership probability for each individual in each cluster, is analogous to STRUCTURE-based membership estimates. We therefore computed values of B, S, and T from the DAPC solutions in the same manner as in the computations with STRUCTURE, propagating the values through analyses parallel to those performed with STRUCTURE-based B, S, and Tand employing the same pipeline. To distinguish DAPC-based measures from STRUCTURE-based measures, we denote DAPC-based measures by B_{DAPC} , S_{DAPC} , and T_{DAPC} .

PCA-STRUCTURE comparisons

PCA-based ancestry information measures produce similar patterns to those observed with STRUCTURE. First, ancestry information is notable with PCA; as in the STRUCTURE analysis, PCA-based inferences are visually similar for the CODIS loci and random non-CODIS sets (Figure S1A-S1D). Measures analogous to *B*, *S*, and *T*, computed using PCA in place of STRUCTURE for the CODIS loci, lie near medians of their respective distributions across random sets (Figure S1E-S1G). The value of T_{DAPC} for PCA, making assignments by applying linear discriminant analysis to PCA coordinates, is 84% (Table S2). Across super-regions, assignment accuracy is greatest for Western Eurasia, the lowest-accuracy super-region for STRUCTURE.

Correlations of PCA-based B_{DAPC} , S_{DAPC} , and T_{DAPC} with \overline{H} , \overline{M} , and $\overline{F_{ST}}$ follow the patterns observed using STRUCTURE-based *B*, *S*, and *T*, with positive values observed between the ancestry information in PCA-based B_{DAPC} , S_{DAPC} , and T_{DAPC} and individual identifiability assessed with \overline{H} and \overline{M} (Table S4). As was seen with STRUCTURE-based *B*, *S*, and *T*, the relationship between information about identity and PCA-based B_{DAPC} , S_{DAPC} , and T_{DAPC} and individual identifiability for STRUCTURE-based and PCA-based population structure analyses is reflected in high correlations between the STRUCTURE-based *B*, *S*, and *T* and the analogous PCA-based B_{DAPC} , S_{DAPC} , and T_{DAPC} (Table S5).

Ancestry studies

The main text notes that among studies at the intersection of forensic genetics and genetic ancestry, some focus on use of statistical measures of marker information content to propose marker panels suitable for ancestry inference, whereas others evaluate the ancestry information for established marker sets such as the CODIS loci. An expanded list of studies in the former class, panel-design, includes [S25-S44]. An expanded list for the latter class, evaluations, includes [S35,S41,S45-S55].

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